



## STUDY ON THE EFFECT OF SPERMINE AND PUTRESCINE ON FREE POLYAMINES AND TSS OF APRICOT FRUITS (VAR: SHAHROODI 48)

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### ABSTRACT

This study was done on Apricot trees (*Prunus armeniaca* var SHAHROODI 48) in Kamal Abad garden in Alborz province. Three concentrations of Put and Spm (5, 10, 15 mM /Lit) was applied on trees in 5 different physiological times (Bud stage, 5 days after Bud stage, Full bloom, 5 days after Full bloom, 10 days after Full bloom). The content of free polyamines (Put, Spm, and Spd) was evaluated. The highest content of Put was observed in Put 15 mM /Lit, Spm in Spm15 mM /Lit, Spd in Spm 15 mM /Lit. The highest content of TSS was observed in Spm 15 mM/Lit.

**Keywords:** apricot, polyamines, spermidine, TSS, bud stage, full bloom.

### INTRODUCTION

Apricot (*Prunus armeniaca* L.) is classified under the *Prunus* species of *Prunodae*, sub-family of the *Rosaceae* and family of the *Rosales* group. Apricot trees can grow over the five continents of the world and production level exceeds 3 million tons (FAO, 2010). Turkey, Iran, Uzbekistan, Italy and Argentina can be regarded as important apricot producer countries. While some of the countries such as Hungary, Morocco, Iran and Tunisia are important fresh apricot exporters, the others, such as Australia and Turkey, are major and famous dried apricot producers and exporters. Dried apricots which are in extensive demand in several parts of the world in USA, UK, Germany, Australia, etc., occupy an important place in the world trade (A.Jannatizadeh *et al.*, 2008). Iran obtain second rate in world production with about 40000 tones and 50000 hectares after Turkey. As known, the fruit of apricot is not only consumed fresh but also used to produce dried apricot, frozen apricot, jam, jelly, marmalade, pulp, juice, nectar, extrusion products, etc.

Polyamines are ubiquitous, aliphatic polycation class of biogenic amines with essential function in living organisms. The most common plant polyamines are the diamine Putrescine and the higher polyamines spermidine and spermine. Spermine and spermidine were early examples of trimethylenediamine derivatives among natural products. Polyamines have been implicated in myriad of biological processes including cell proliferation, cell division and differentiation, apoptosis, homeostasis, gene expression, protein and DNA synthesis. In plants, Putrescine is formed from either argentine via an intermediate agmatine, a reaction catalyze by arginine decarboxylase, or from ornitine by ornitine decarboxylase. Spermidine is synthesized from Putrescine and the aminopropyl group donated by decarboxylated S-

adenosylmethionine (SMA), which is a product of SMA decarboxylation (Figure-1) (Autar K.Mattoo *et al.*, 2008).

Putrescine is known to improve plant growth and development due its effects on cell division and differentiation. Such findings were confirmed in bean plants by Altman *et al.*, in 1982. Moreover, Putrescine (as one of the polyamine group) has a regulatory role in promoting productivity of many plants such as sweet pepper (Talaat., 2003), tomato (Cohen *et al.*, 1982) and pea plants by Gharib and Hanafy, 2005 (El-Tohamy *et al.*, 2008).

### MATERIAL AND METHODS

This experiment was done in Apricot collection in Kamal Abad Garden which located in Kamal Abad city, Karaj, Alborz province. Traditional variety (SHAHRODI 48) was selected for this study. This Variety is one of the early seasons of apricot trees. After preparing 5, 10 and 15 mM/Lit of Spermine and Putrescine, they are painted on buds in two different times, bud stage and 5 days after bud stage. Because of the sensitivity of blooms, we have to spray solutions on blooms in three different times, Full Bloom, 5days after full bloom and 10 days after full bloom. Sampling was done in three stages; small fruit stage, peat hardening stage and full fruit set.

Polyamines were evaluated with HPLC and with Torrigiani method. In the laboratory TSS was valued with Refractrometer.

### RESULT AND DISCUSSIONS

The result of analysis of variance Table-1, shown that the content of free polyamines were affected with Spm, and Put. This content has a significant effect on the level of 1 %. According to the table using Polyamines had no significant effect on the TSS (Table-1).



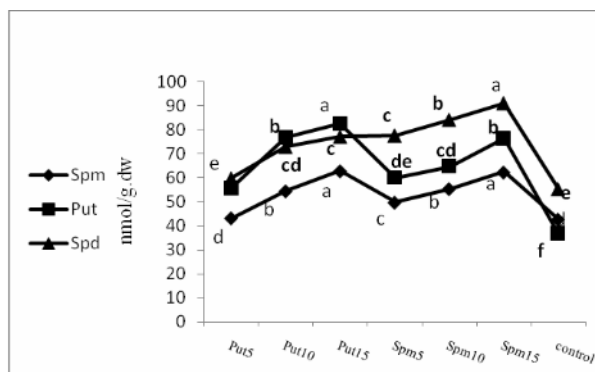
**Table-1.** The effect of polyamines treatment on free polyamines and content of TSS of fruit of Apricot Var. SHAHROODI 48.

Source	FD	TSS	Spd	Put	Spm
Stage	4	171.02**	42406.33**	65758.51**	30020.89**
Time	3	733.29**	86952.96**	78449.68**	28307.83**
Treatment	9	6.20 <sup>ns</sup>	6141.36**	8120.60**	14094.99**
S*T	12	31.37**	12540.88**	12075.96**	5601.23**
S*Tr	36	3.46**	638.80**	1346.11**	518.73**
T*Tr	27	21.40**	4494.12**	2989.27**	3415.30**

\*\*,\*and ns: signification on the level of 1%, 5% and non significant

**The effects of treatment on free Spm, Spd and Put**

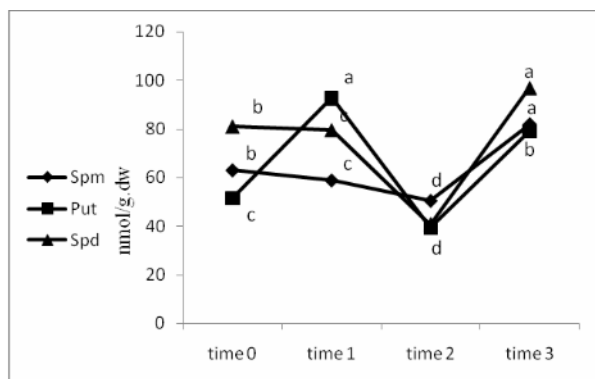
According to Figure-1 the highest content of Spm and Put was observed in Put1 5. The lower content of Spm and Put obtained in control samples. The highest content of free Spd obtained in Spm 15 and the lowest content of it obtained in Put 5 and control samples.



**Figure-1.** The effect of treatment on free Spm, Put, Spd.

**The effects of time on free Spm, Put, and Spd**

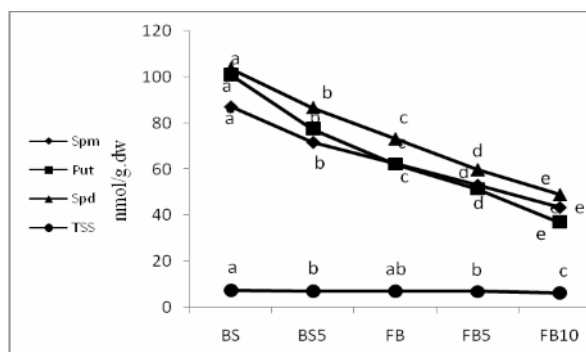
As it shows in Figure-2, the content of free polyamines was affected by time. The highest content of Spm and Spd was obtained in time 3. The highest content of Put was obtained in time 1. The lowest content of these polyamines were observed in time 2.



**Figure-2.** The effect of time on free Spm, Put, Spd.

**The effects of time on free Spm, Put, Spd and TSS**

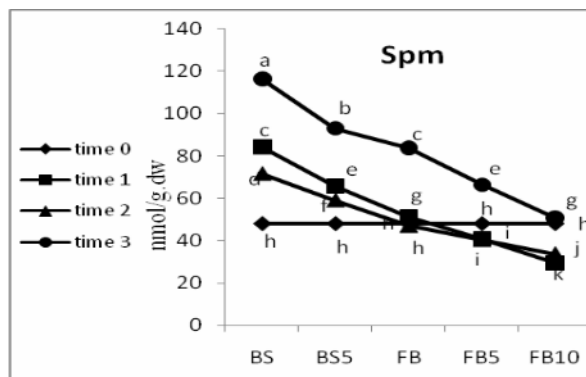
According to Figure-3, the content of free polyamines was affected by Spm and Put. The highest content was observed in BS and the lowest was obtained in FB10. But the effect of these polyamines on the percentage of TSS hadn't significant effect between BS5 and FB5. So in BS it has the highest percentage and in FB10 had the lowest percentage.



**Figure-3.** The effect of stage on free Spm, Put, Spd, TSS.

**The effects of time and stage interactions on free Spm**

According to Figure-4, the highest content of Spm was observed in BS and time 3. The lowest content of Spm was obtained in FB10 and time 1.

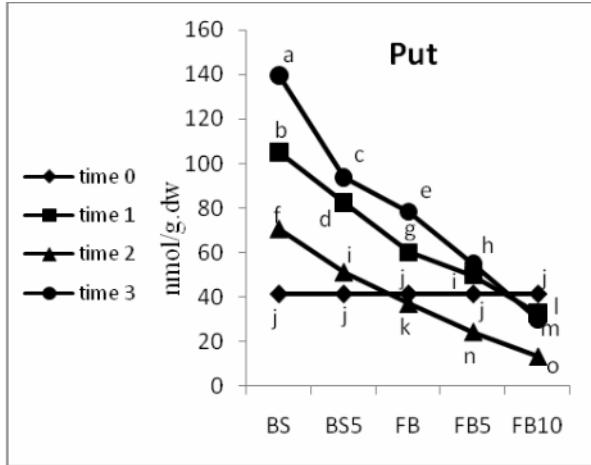


**Figure-4.** The effects of time and stage interactions on free Spm.



**The effects of time and stage interactions on free Put**

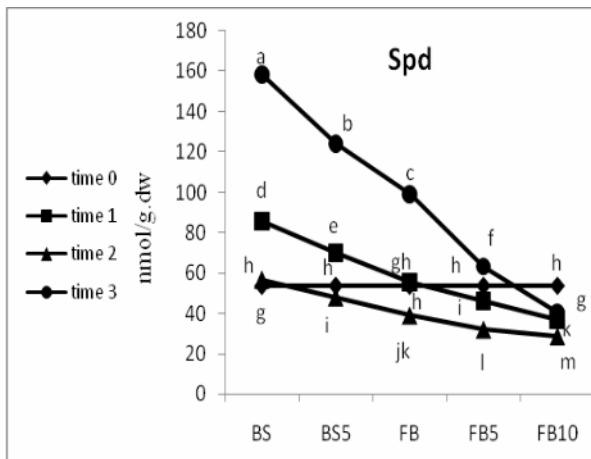
According to Figure-5, the highest content of Put was observed in BS and time 3. The lowest content of Put was obtained in FB10 and time 2.



**Figure-5.** The effects of time and stage interactions on free Put.

**The effects of time and stage interactions on free Spd**

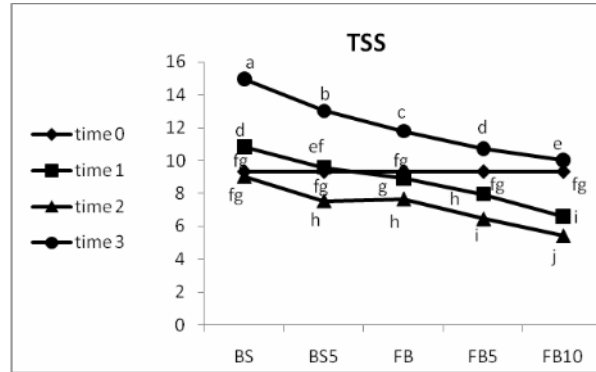
According to Figure-6, the highest content of Spd was observed in BS and time 3. The lowest content of Spd was obtained in FB10 and time 2.



**Figure-6.** The effects of time and stage interactions on free Spd.

**The effects of time and stage interactions on TSS**

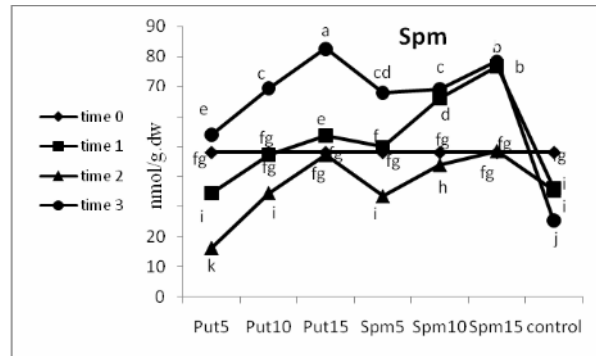
According to Figure-7, the highest percentage of TSS was observed in BS and time 3. The lowest percentage of TSS was obtained in FB10 and time 2.



**Figure-7.** The effects of time and stage interactions on TSS.

**The effects of time and treatment interactions on free Spm**

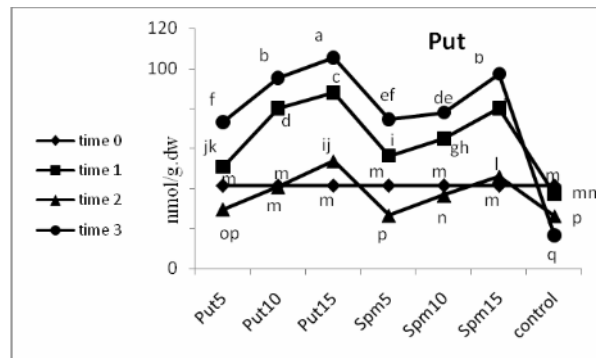
According to Figure-8, the highest content of Spm was observed in Put 15 and time 3. The lowest content of Spm was obtained in Put 5 and time 2.



**Figure-8.** The effects of time and treatment interactions on Spm.

**The effects of time and treatment interactions on free Put**

According to Figure-9, the highest content of Put was observed in Put 15 and time 3. The lowest content of Put was obtained in Put5 and time 2.

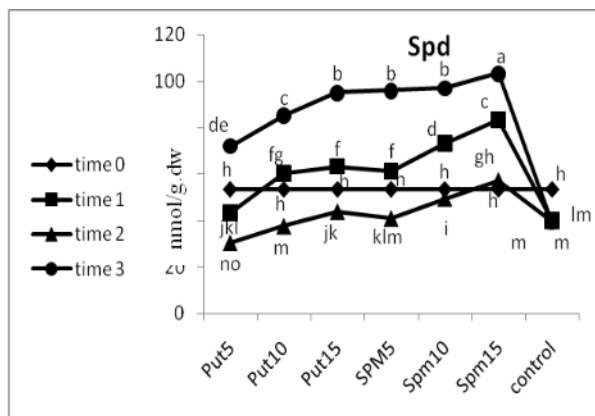


**Figure-9.** The effects of time and treatment interactions on Put.



**The effects of time and treatment interactions on free Spd**

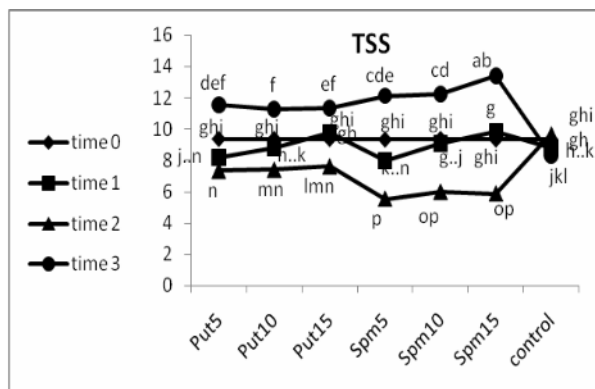
According to Figure-10, the highest content of Spd was observed in Spm 15 and time 3. The lowest content of Spd was obtained in Put5 and time 2.



**Figure-10.** The effects of time and treatment interactions on Spd.

**The effects of time and treatment interactions on TSS**

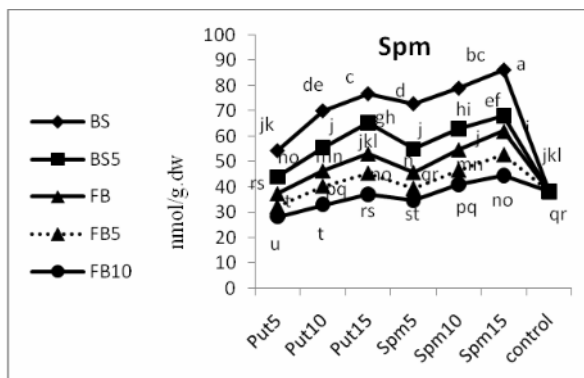
According to Figure-11, the highest percentage of TSS was observed in Time 0 and Spm15. The lowest percentage of TSS was obtained in Spm5 and time 2.



**Figure-11.** The effects of time and treatment interactions on TSS.

**The effects of stage and treatment interactions on Free Spm**

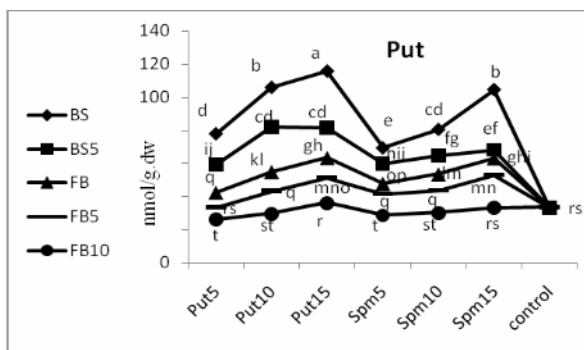
According to Figure-12, the highest content of Spm was observed in BS and Spm 15. The lowest content of Spm was obtained in FB10 and Put 5.



**Figure-12.** The effects of stage and treatment interactions on Spm.

**The effects of stage and treatment interactions on free Put**

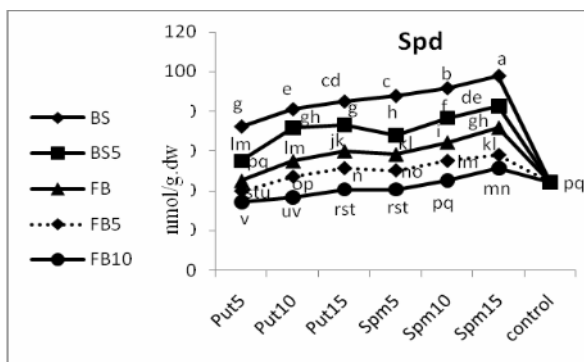
According to Figure-13, the highest content of Put was observed in BS and Put 15. The lowest content of Put was obtained in FB10 and Spm5.



**Figure-13.** The effects of stage and treatment interactions on Put.

**The effects of stage and treatment interactions on free Spd**

According to Figure-14, the highest content of Spd was observed in BS and Spm 15. The lowest content of Spd was obtained in FB10 and Put 5.

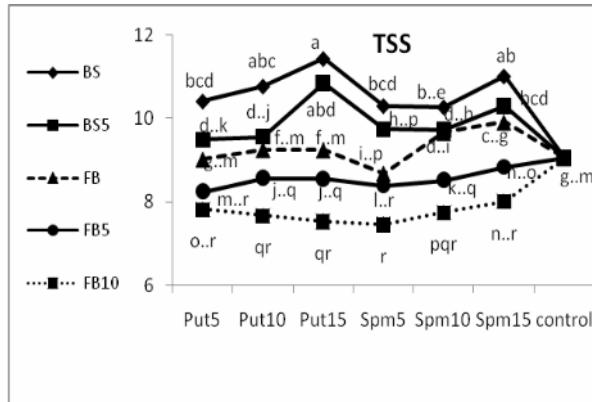


**Figure-14.** The effects of stage and treatment interactions on Spd.



### The effects of stage and treatment interactions on TSS

According to Figure-15, the highest content of TSS was observed in BS and Put 15. The lowest content of TSS was obtained in FB10 and Spm5.



**Figure-15.** The effects of stage and treatment interactions on TSS.

The general positive effects observed at this study on fruit characteristics of apricot fruits as a result of applying polyamines could be attributed to enhancement effects of polyamines applications on floral developments which resulted on endogenous polyamines. In addition, accumulating evidences has showed that initial concentration of Put, Spm, Spd, and total free polyamines in the pulp and seeds were a little high for both Zoami pears and Whang Keumbea pears. It might be induced by vigorous activities of cell division in fruit at the early stage (Xu Liu *et al.*, 2008). Aussanee Paksasorn *et al.* (1995) reported that polyamines have been shown to be associated with cell division during early developments, usually increasing when rapid cell division occur. In free PAs, especially in the dominant Put the highest level was found at Full Bloom in all three cultivars, as in other plants (Shuji Shiozaki *et al.*, 2002). In cultivars with high PA content in the early phase of fruit growth the fruit cell division process might be more active (Omar Franco-Moral *et al.*, 2005). Spd application increases other endogenous polyamines content (Put, Spm) or (Put, Spm, Spd) in cotton leaves at both vegetative and flowering stages. The results indicate that PAs content were more enhanced in the leaves at flowering stage than vegetative stage (M.S.A. Abd El-Wahed, 2006). Canino apricot trees grown under sandy soil conditions greatly respond to foliar spraying with polyamines to enhance production and improve fruit quality (Enas A.M. Ali *et al.*, 2010).

### CONCLUSIONS

In conclusion, this study showed that the content of free polyamines and TSS illuminated with exogenous polyamines. These results suggest that exogenous polyamines may be an effective way for increase or decrease free polyamines which may effect on flowering and ripening. So the best treatment for increasing the

content of Put, Spm and Spd was Put 15 mM /Lit, Spm15 mM /Lit, and Spm 15 mM /Lit. The best treatment for increasing the content of TSS was Spm 15 mM/Lit. Further investigation is needed to study the effect of them on flowering trees. Fruits of Apricot trees treated with both (Put) and (Spm) had a significantly higher soluble solids concentration (SSC) and it was firmer than fruits of control tree. The best time for treat Apricot trees in at Bud Stage for increasing polyamines.

### REFERENCES

- Akiva Apelbanum, Alan Brurgoon, James D. Anderson and Morris Lieberman. 1981. Polyamines Inhibit Biosynthesis of Ethylene in Higher Plant Tissue and Fruit Protoplasts. *Plant Physiol.* 68: 453-456.
- Anna M. Bregoli *et al.* 2002. Peach (*Prunus persica*) fruit ripening: aminoethoxyvinylglycine (AVG) and exogenous polyamines affect ethylene emission and flesh firmness. *Physiologia Plantarum.* 114(3): 472-481.
- Anne Santerre *et al.* 1990. Effect of acid rain on polyamines in Picea. *Phytochemistry.* 29(6): 1767-1769.
- Asghari and Khamiri Sani. 2010. Effect of post harvest application of nitric oxide and Putrescine on some qualitative properties and phenolic content of the white grape varieties. *Food Research Journal.* 3(2).
- Aussanee Paksasorn, Tomoko Hayasaka, Hiroyuki Matsui, Hitoshi Ohara and Naomi Hirata. 1995. Relationship of Polyamine Content to ACC Content and Ethylene Evolution in Japanese Apricot Fruit. *J. Japan. Soc. Hort. Sci.* 63: 761-766.
- Autr K. Mattoo, Avtr K. H and. 2008. Higher polyamines restore and enhance metabolic memory in ripening fruit. *Plant science.* 174: 386-393.
- Bueno M. and Matilla A. 1992. Abscisic acid increases the content of free polyamines and delays mitotic activity induced by spermine in isolated embryonic axes of chickpea seeds. *Physiol. Plant.* 85: 531-536.
- Cesar Petri *et al.* 1992. Spermidine pretreatment of root tip removal in maize seedlings: effects on K<sup>+</sup> uptake and tissue modifications. *J. Plant Physiol.* 140: 741-746.
- Ching Hwei Kao. 1993. Polyamines and rice leaf senescence. *Bot. Bull. Acad. sin.* 34: 299-305.
- David G. Davis and Prudence A. Olson. 1994. Effects of Putrescine and inhibitors of Putrescine biosynthesis on organogenesis in *Euphorbia esula* L. in Vitro Cellular and Developmental Biology - Plant. 30(2): 124-130.
- El-Tohamy W.A., H.M. El-Abagy and N.H.M. El-Greadly. 2008. Studies on the Effect of Putrescine, Yeast





and vitamin C on Growth, Yield and Physiological Responses of Eggplant (*Solanum melongena* L.) Under Sandy Soil Conditions. Australian Journal of Basic and Applied Sciences. 2: 296-300.

Enas A.M Ali *et al.* 2010. Improving Canino Apricot Trees Productivity by Foliar Spraying with Polyamines. Journal of Applied Sciences Research. 6(9): 1359-1365.

greenacresblog.files.wordpress.com.

H.J. Scholten. 1998. Effect of polyamines on the growth and development of some horticultural crops in micro propagation. Scientia Horticulturae. 77(1-2): 83-88.

Haji boland and Ebrahimi. 2011. Effect of exogenous polyamines on growth, photosynthesis and metabolism of phenols in tobacco plants under salt stress. Plant Biology, Third Year. 8: 13-26.

Harsh Pal Bais and G.A. Ravishankar. 2000. Influence of polyamines on growth and formation of secondary metabolites in hairy root cultures of *Beta vulgaris* and *Tagetes patula*. Acta Physiologiae Plantarum. 22(2): 151-158.

Jihong Liu, Kazuyoshi Nada, Xiaoming Pang, Chikako Honda, Hiroyasu Kitashiba and Takaya Moriguchi. 2006. Role of polyamines in Peach Fruit Development and Storage. Tree Physiology. 26: 791-798.

Kursat Cavusoglu and Kudret Kabar. 2007. Comparative effects of some plant growth Regulators on the germination of barley and radish seeds under high temperature stress. EurAsia J. BioSci. pp. 1-10.

LI Jian-yong *et al.* 2005. Advances in research on effects of polyamines on fruit growth and development of fruit trees. Journal of Fruit Science. 2005-03.

M.S.A. Abd El-Wahed. 2006. Exogenous and Endogenous Polyamines Relation to Growth,  $\alpha$ -cellulose Precipitation in Fibres and productivity of Cotton Plant. World Journal of Agricultural Sciences. 2: 139-148.

M.S.A. Abd EL-Wahed. 2006. Exogenous and Endogenous Polyamines Relation to Growth,  $\alpha$ -cellulose Precipitation in Fibres and Productivity of Cotton Plant. World Journal of Agricultural Sciences. 2(2): 139-148.

Marianthi H. Loupassaki, Ioannis I. Androulakis and Miltiadis D. Vasilakakis. 1995. Effect of Polyamines, Gibberellins and Other Growth Regulators on the Fruit-Set of Avocado. Proceedings of the World Avocado Congress 3: 57-60.

Marie-Claire Heloir, Clair Kevers, Jean-Francois Hausman and Thomas Gaspar. 1996. Changes in the concentrations

of auxins and polyamines during rooting of in-vitro-propagated Walnut shoots. Tree Physiology. 16: 515-519.

Mark A. Walker, Dane R. Roberts, Ching Yu Shih and Erwin B. Dumbroff. 1985. A Requirement for Polyamines during the cell Division Phase of Radicle Emergence in Seeds of *Acer saccharum*. Plant and cell Physiology. 26(5): 967-971.

Mercedes Ariasa, Juan Carbonellb and Manuel Agustí. 2005. Endogenous free polyamines and their role in fruit set of low and high parthenocarpic ability citrus cultivars. Journal of Plant Physiology. 162(8): 845-853.

Miguel A. Perez-Amador and Juan Carbonell. 1995. Arginine Decarboxylase and Putrescine Oxidase in Ovaries of *Pisum sativum* L. Plant Physiol. 107: 865-872.

Moghim and *et al.* 2010. Protective role of polyamines against heat shock on growth of soybean seedlings. IRAN Plant Biology, Second year, 2<sup>nd</sup> Ed. (s 4): 31-40.

Mosbah M. Kushad, George Yelenosky and Robert Knight. 1988. Interrelationship of Polyamine and Ethylene Biosynthesis during Avocado fruit Development and ripening. plant Physiol. 87: 463-467.

Muhammad Muzammil Jahangir *et al.* 2011. Effect of Spermine on Bioactive Components and Antioxidant Properties of Sliced Button Mushroom (*Agaricus bisporus*) during Storage. International Journal of Agriculture and Biology Issn Print: 1560-8530; ISSN Online: 1814-9596.

N. Béranger Novat *et al.* 1994. Polyamines and their biosynthetic enzymes in dormant embryos of the spindle tree (*Euonymus europaeus* L.) and in dormancy break obtained after treatment with gibberellic acid. Plant Science. 102(2): 139-145.

N. Savithramma and P. M. Swamy. 1989. Quantitative Changes of Polyamines and Activity of ADC in Developing and Germinating Seeds of Two Groundnut (*Arachis hypogaea*) Genotypes, Annals of Botany. 64: 337-341.

Naoko Wada, Masateru Shinozaki and Hajime Iwamura. 1994. Flower Induction by Polyamines and Related Compounds in Seedlings of Morning Glory (*Pharbitis nil* cv. Kidachi). Plant and Cell Physiology. 35(3): 469-472.

Omar Franco-Mora, Kenji Tanabe, Akihiro Itai, Fumio Tamura and Hiroyuki Itamura. 2005. Relationship between Endogenous Free Polyamine Content and Ethylene Evolution during Fruit Growth and Ripening of Japanese Pear (*Pyrus pyrifolia* Nakai). J. Jpn. Soc. Hort. Sci. 74: 221-227.



Philip Coffino. 2001. Biosynthesis and catabolism of polyamines. *Nature Reviews Molecular Cell Biology*. 2: 188-194.

Putrescine treatments on in vitro rooting and on endogenous polyamine changes in difficult-to root woody species. *Scientia Horticulturæ*. Elsevier Science Publishers B.V., Amsterdam. 53: 63-72.

R.K. Kakkar *et al.* 2000. Polyamines and Plant Morphogenesis. *Biologia Plantarum*. 43(1): 1-11.

Ravindar Kaur-Sawhney *et al.* 2003. Polyamines in plants: An overview. *Journal of Cell and Molecular Biology*. 2: 1-12.

Rugini Eddo *et al.* 1993. Role of basal shoot darkening and exogenous.

S. Scaramagli, M. Francescetti, A.J. Michael, P. Torrigiani and N. Bagni. 1999. Polyamines and flowering: spermidine biosynthesis in the different whorls of developing flowers of *Nicotiana tabacum* L. *Plant Biosystem*. 1333: 229-237.

Saif and *et al.* 2008. Effects of polyamines and benzyladenine treatments to preserve the nutritional properties of pomegranate (*Punica granatum* L. cv. Rabbab) during mass storage, Eighteenth Congress of Food Science and Technology.

Shuji Shiozaki, Tsuneo Ogata and Shosaku Horiuchi. 2002. Differences in Polyamine Levels between "Kyoho", "Campbell Early" and "Muscata Bailey A" Grapes with differing Features of Berry Drop. *Sci. Rep. Agric. And Biol. Sci., Osaka Pref. Univ.* 54: 15-23.

T.A. Smith. 1970. Putrescine, spermidine and spermine in higher plants, *Phytochemistry*. 9(7): 1479-1486.

Talaih and *et al.* 2010. Study on Effect of spraying Polyamines on some physiological problems in Pistachio var Kaleghoochii. *Journal of Horticultural Science*. 41(4): 383-391.

Theodora S. Pritsa and Demetrios G. Voyiatzis. 2005. Correlation of ovary and leaf spermidine and spermine content with the alternate bearing habit of olive. *Journal of Plant Physiology*. 162(11): 1284-1291.

Ural Meral, Narcin Palavan-Unsal and Isil Ismailoglu. 2005. The Effect of Putrescine and difluoromethylornitine on cell division activity of wheat in different ploidy level. *caryologia*. 58(1): 15-20.

Wie Tang and Ronald J. Newton. 2005. Polyamines promote root elongation and growth by increasing root cell division in regenerated Virginia pine (*Pinus virginiana* Mill.) plantlets. *Plant Cell Reports*. 24(10): 581-589.

www.wikipedia.Com.

Xu Liu, Ming-an Liao, Guo-tao Deng, Shan-bo Chen, Ya-jun Ren and Wei-Guo Liu. 2008. Changes in Endogenous Hormones and Polyamines of fruit During Growth and Development of Pear Fruits. *World Journal of Agricultural sciences*. 4: 40-47.

Z. Singh and J. Janes. Regulation of Fruit Set and Retention in Mango with Exogenous Application of Polyamines and Their Biosynthesis Inhibitors. *Ishs Acta Horticulturæ*. p. 509.